# MEDICAL ONCOLOGY



# Biomarkers that currently affect clinical practice: *EGFR, ALK, MET, KRAS*

M.D. Vincent MB ChB,\* M.S. Kuruvilla MD,\* N.B. Leighl BSc MD MSc,† and S. Kamel–Reid PhD†

# **ABSTRACT**

New drugs such as pemetrexed, the epidermal growth factor receptor (EGFR) tyrosine kinase inhibitors, and the Alk inhibitor crizotinib have recently enabled progress in the management of advanced non-small-cell lung cancer (NSCLC). More drugs, especially Met inhibitors, will follow. However, the benefits of these agents are not uniform across the spectrum of NSCLC, and optimizing their utility requires some degree of subgrouping of NSCLC by the presence or absence of certain biomarkers.

The biomarkers of current or imminent value are *EGFR* and *KRAS* mutational status, *ALK* rearrangements, and *MET* immunohistochemistry. As a predictor of benefit for anti-EGFR monoclonal antibodies, *EGFR* immunohistochemistry is also of potential interest.

Some of the foregoing biomarkers (*EGFR*, *ALK*, *MET*) are direct drivers of the malignant phenotype. As such, they are, quite rationally, the direct targets of inhibitory drugs. However, *KRAS*, while definitely a driver, has resisted attempts at direct pharmacologic manipulation, and its main value might lie in its role as part of an efficient testing algorithm, because *KRAS* mutations appear to exclude *EGFR* and *ALK* mutations. The indirect value of *KRAS* in determining sensitivity to other targeted agents or to pemetrexed remains controversial. The other biomarkers (*EGFR*, *ALK*, *MET*) may also have indirect value as predictors of sensitivity to chemotherapy in general, to pemetrexed specifically, and to radiotherapy and molecularly targeted agents.

These biomarkers have all enabled the co-development of new drugs with companion diagnostics, and they illustrate the paradigm that will govern progress in oncology in the immediate future. However, in NSCLC, the acquisition of sufficient biopsy material remains a stubborn obstacle to the evolution of novel targeted therapies.

# **KEY WORDS**

NSCLC, lung cancer, EGFR, ALK, KRAS, prognosis, prediction

#### 1. INTRODUCTION

Metastatic non-small-cell lung cancer (NSCLC) remains, with rare surgical exceptions, incurable. Pending radical new solutions, scientific progress is currently channelled into the conversion of this rapidly lethal disease into a chronic illness. How to make that conversion is conventionally believed to involve "personalized medicine": Tumour biopsies are tested for certain causative and characteristic molecular lesions ("targets"), guiding the selection of customized drugs designed to directly interact with and inhibit those targets.

This paradigm, based on the concept of causality, is deeply seductive, given that it appears to offer the prospect of both efficacy and lack of toxicity. It hews to a fundamentally rational worldview as suggested by its common appellation, "targeted therapy." The molecular lesion is meant to be causally responsible for maintenance of the malignant phenotype and also distinctive, even uniquely characteristic, of the cancer cells. Hence the prospects for both tumour control and selectivity.

The foregoing perspective, while undoubtedly simplistic, nonetheless provides a framework for how four key genes—*EGFR*, *ALK*, *MET*, and *KRAS*—will increasingly influence the management of metastatic NSCLC. Those genes, when altered in measurable ways, unquestionably contribute to the pathogenesis of NSCLC, and as such, are widely agreed to be "drivers" <sup>1</sup>. It is rational to seek to inhibit them, even to hope that drugs can be designed that will selectively block the oncogenic varieties while sparing their normal counterparts. But there is no guarantee that oncogenic variants are necessarily druggable, or if they are, that the cancer cell will not eventually find a way around

the inhibition. Furthermore, those genes, although not necessarily mutated *sensu stricto*, may, through altered expression, nonetheless still contribute to the cancer, thus reducing the prospects for selectivity.

Additionally, genetic aberrations such as these may convey useful information beyond the notion of the direct target. Broadly, that information can be classified as prognostic (foreknowledge of probable events in the absence of therapy, which may continue to influence outcomes regardless of therapy) and predictive (indicating the prospects for success of particular therapies), and might be mechanistically related to the aberrant gene, but might also be purely empirical—that is, exhibiting no obvious causal relationship. These genetic alterations, then, are "biomarkers" sensu lato, and their utility extends into predicting the future clinical course (even absent therapy) and the selection of drugs (whether those drugs target those particular genes directly or not).

It is better, therefore, to approach *EGFR*, *ALK*, *MET*, and *KRAS* and the entire expanding suite of molecular drivers <sup>2</sup> as biomarkers in the broad sense, and not just as direct targets, although the latter status is clearly of major importance.

Although the present review focuses more on the biomarker utility of these genes and less on the technicalities of their measurement, we must emphasize that the acquisition of adequate biopsy material remains problematic in the management of metastatic NSCLC. That problem can partly be addressed by educating respirologists, interventional radiologists, and thoracic surgeons, but sometimes there is no possibility of obtaining other than scant tissue. The reasons include hazard, technical factors, access, patient refusal, and avoidance of delay.

In the event that the clinician's hand is forced, we therefore provide information correlating the foregoing biomarkers with (usually available) clinical, pathologic, and demographic characteristics. Emphatically, however, it is better to make therapeutic decisions on the basis of a direct test. However, as a definitive solution to this problem, reliable testing based on blood work (that is, analysis of circulating tumour cells or plasma DNA) should soon become available <sup>3,4</sup>.

# 2. EGFR

In the early 1960s, Stanley Cohen isolated the mitogen "epidermal growth factor" (EGF) from murine salivary gland <sup>5</sup>. In 1973, the EGF receptor (EGFR) was described <sup>6</sup>; this receptor was later appreciated as the first of a family of 4 human epidermal tyrosine kinase receptors (HER1–4) <sup>7</sup>, attended by a broad spectrum of ligands besides EGF, participating in a multifaceted and adaptive signalling network <sup>8</sup> subserving growth and survival. *EGFR*, cloned and isolated in 1984 <sup>9</sup>, encodes a 1210-amino-acid transmembrane protein, including an extracellular ligand-binding ectodomain, an anchoring transmembrane domain, and a submembrane

tyrosine kinase domain. Ligand activation involves homo-dimerization (or hetero-dimerization with other HER family members), and then activation of the tyrosine kinase domain, resulting in tyrosine autophosphorylation, which enables engagement with 6 or more signalling pathways subserving "cell fate decisions", including the PI3K/Akt and Erk pathways of particular interest in oncology.

Dysregulation of EGFR contributes to a range of cancers and occurs in various ways <sup>10</sup>. In NSCLC, the most important are activating *EGFR* mutations and increased protein expression. Either dysregulation may possibly be associated with increased gene copy number. The uncommon *EGFRvIII* mutation has also been detected in a few squamous cell lung cancers <sup>1</sup>. However it arises, dysregulated *EGFR* activation promotes the malignant phenotype by mediating cell proliferation, raising the apoptotic threshold, increasing cellular motility (and hence metastasis), enhancing neoangiogenesis, and conferring resistance to chemotherapy and radiation.

Although earlier efforts at predicting anti-EGFR therapeutic sensitivity focused on EGFR protein overexpression and EGFR gene copy number increment, the most important parameter is whether an activating EGFR mutation is present. The mutations are almost exclusively found in lung adenocarcinomas; they are more common in never-smokers or light exsmokers, women, and patients of East Asian origin. In this demographic, 60%–70% of patients will have a detectable mutation in EGFR. Caucasian smokers or ex-smokers with adenocarcinomas have an 8% incidence—enough to mandate testing. All patients with adenocarcinomas should be tested for EGFR mutation (Table 1) 11-13, although that dictum may need to be softened depending on immunophenotyping. Mutations are associated mainly with papillary and micropapillary adenocarcinomas or non-mucinous bronchioloalveolar adenocarcinomas (rarely with solid adenocarcinomas) and seem mostly to require an immunophenotype positive for thyroid transcription factor 1 (TTF-1).

Nearly all activating *EGFR* mutations occur in exons 18–21. The most important are deletions within exon 19 (more than 20 variants) and point (missense) mutations in exon 21 (usually L858R,

TABLE I Estimated genomic probabilities in adenocarcinoma 11-13

Variable	Value by locale			
	East Asia <sup>a</sup>	Western world $^b$		
Studies (n)	6	2		
Patients (n)	814	116		
Never-smokers, EGFR M+ (%)	70	37		
Ever-smokers, EGFR M+ (%)	29	8		

a Japan, Korea, Taiwan, Hong Kong.

b United States, Australia.

occasionally L861Q or L861R). Very occasionally, point mutations involve exon 18 (for example, G719C and others at G719). Generally the tyrosine kinase domain is affected, probably leading to increased ATP binding, with enhanced (and ligand-independent) downstream signalling, especially via the Akt and STAT pathways, affecting cell survival <sup>14</sup>. The resulting condition ("oncogene addiction") is characterized by a dependency of the cancer cell on the mutation. Also implicated is the Erk1/2 pathway, essential to cellular proliferation <sup>15</sup>. The benefits of EGFR blockade may ultimately be mediated by a shift toward apoptosis in the balance of the pro- and anti-apoptotic members of the Bcl-2 family of proteins.

The centrality of EGFR signalling has led to intensive efforts to design therapies aimed at blockade. Two approaches have proved successful: anti-EGFR monoclonal antibodies against the extracellular ligand-binding domain, and small-molecule tyrosine kinase inhibitors (TKIS) to block binding of ATP (upon which signalling depends). The latter have proved much more valuable in NSCLC, although EGFR antibodies have also demonstrated activity.

Curiously, small-molecule TKIS (gefitinib and erlotinib) were designed before the elucidation, in 2004 by three American groups, of the EGFR mutation <sup>2,16,17</sup>. The small subset of metastatic NSCLC patients who had responded dramatically to singleagent TKI therapy prompted the search for an explanation, culminating in discovery of the mutations. These mutations not only confer oncogene addiction, but also fortuitously show markedly increased affinity for gefitinib or erlotinib because of residue repositioning around the binding cleft <sup>18</sup>. It soon became apparent that almost all the dramatic responses had occurred in patients whose cancers harboured one of these activating (and sensitizing) EGFR mutations; however, EGFR-TKI can also, to a lesser extent, benefit patients without those mutations: that is, the EGFR "wild-type" (EGFR WT) patients, whose cancers are presumably driven by upregulated signalling (from overexpression of the normal protein, for instance).

Small (mainly East Asian) studies of EGFR-TKI monotherapy with gefitinib rapidly confirmed high objective response rates (55%–91%) in patients with cancers harbouring a mutation 19-26. A large nonrandomized 217-patient Spanish-led experience <sup>27</sup> with erlotinib was published in 2009. The objective response rate (ORR) of 70.6%, the progressive disease rate of just 10.2%, the prolonged progression-free survival (PFS) of 14 months, and the overall survival (os) of 27 months suggested that responsiveness in mutationpositive patients was not a function of ethnicity and that erlotinib might be superior to gefitinib. Furthermore, Caucasian patients demonstrated a spectrum of EGFR mutational subtypes similar to those seen in East Asian patients. Those phase II trials led to six large randomized trials comparing first-line EGFR-TKI with then-standard platinum-doublet third-generation

chemotherapy in proven *EGFR* mutation–positive patients (*EGFR* M+) or in populations enriched for mutation positivity (Table II).

The randomized studies (IPASS, WJTOG 3405, NEJ002, First-signal, optimal, and Eurtac) uniformly revealed that, compared with chemotherapy, first-line TKI consistently resulted in a higher ORR and longer PFS; however, os was not prolonged because of extensive crossover from chemotherapy to TKI upon progression. Because TKI and chemotherapy appear non-cross-resistant, those who receive a second-line TKI benefit as much as those who receive it in the first line <sup>27,35</sup>. However, because of unavoidable attrition (35% in IPASS), it is desirable to treat with a TKI up front if possible in EGFR M+ patients, notwithstanding a modest delay to secure a test result. However, as revealed by IPASS, the one trial to accrue and analyze both EGFR M+ and WT patients, the opposite is even more true. Clearly, in EGFR WT disease (approximately 40% of the East Asian IPASS population of never-smokers or light ex-smokers), gefitinib appears virtually devoid of useful activity (ORR: 1.1%) and may be associated with passive harm because of the opportunity cost of delaying active chemotherapy.

In the trials, patients with exon 19 deletions and exon 21 point mutations did not have markedly different outcomes on TKI (the former perhaps conferring a modestly better outcome). Also, erlotinib is probably not markedly different from gefitinib in outcome, and (from IPASS) EGFR mutation positivity is prognostic for inherently longer survival. There is a suggestion (again from IPASS) that, compared with EGFR WT patients, those who are EGFR M+ respond better to chemotherapy, although the ORR in IPASS for M+ patients (47%) was outside the range for chemotherapy in the other five randomized trials (15%–37%). However, IPASS did establish that firstline chemotherapy in EGFR M+ patients was much more active than TKI in EGFR WT patients, implying that EGFR-unknown patients should receive first-line chemotherapy rather than "empirical TKI."

A subsequent randomized trial of post-chemotherapy maintenance erlotinib compared with placebo (SATURN) exhibited a dramatic benefit for the *EGFR* M+ subset [hazard ratio (HR): 0.10] in PFS, but not in os, again because of crossover. Interestingly, the *EGFR* WT patients did experience an os advantage—but only if the best response on prior first-line chemotherapy was stable disease, not complete or partial response <sup>36</sup>.

Mutations in *EGFR* also occur in exon 20, especially T790M, which inserts a bulky methionine over the ATP binding cleft, blocking access to first-generation EGFR-TKI (but not to ATP) <sup>37</sup>. This "gatekeeper" T790M mutation occurs only within a pre-existing sensitizing mutation, either del 19 or exon 21, and seemingly causes up to 50% of the resistance inevitably occurring in all *EGFR* M+ patients on first-generation TKI (gefitinib or erlotinib). Novel EGFR-TKI (for example, afatinib, dacomitinib)

#### BIOMARKERS AFFECTING CLINICAL PRACTICE

TABLE II Randomized trials of chemotherapy compared with epidermal growth factor receptor (EGFR) tyrosine kinase inhibitor (TKI) in mutation-positive patients

Reference (study name)	Regimen	ORR (%)	Statistic	p Value	PFS (months)	HR	p Value	OS (months)	HR	p Value
Lee et al., 2009 <sup>28</sup> ; Ku et al., 2011 <sup>29</sup>	Cisplatin–gemcitabine vs.	38		0.002	6.7		0.0084	26.5	HR?	0.648
(First-signal)	gefitinib	85			8.4			30.6		
Maemondo <i>et al.</i> , 2010 <sup>30</sup> (NEJ 002)	Carboplatin–paclitaxel vs.	31		< 0.001	5.4	0.30	< 0.001	23.6	Not avail-	0.31
	gefitinib	74			10.8			30.5	able	
Mitsudomi <i>et al.</i> , 2010 <sup>31</sup> (wJTOG 3405)	Cisplatin-docetaxel	32		< 0.001	6.3	0.49	<0.0001	Not	1.64	0.211
	vs. gefitinib	62			9.2			reached	30.9	
Fukuoka <i>et al.</i> , 2011 <sup>32</sup> ; Mok, 2011 <sup>a</sup> (IPASS)	Carboplatin-paclitaxel	47		< 0.001	6.3	0.48	< 0.001	21.9	1.0	0.99
	vs. gefitinib	71			9.5			21.6		
Zhou <i>et al.</i> , 2011 <sup>33</sup> (OPTIMAL)	Carboplatin– gemcitabine vs.	36		<0.0001	4.6	0.16	<0.0001	Not avail- able	Not avail- able	
	erlotinib	83			13.1			uoic	uoie	
Rossell <i>et al.</i> , 2012 <sup>34</sup> (EURTAC)	Platinum–gemcitabine or platinum–docetaxel vs.	15	or: 7.5	<0.0001	5.2	0.37	<0.0001	19.5	1.047	0.87
	erlotinib	58			9.7			19.3		

<sup>&</sup>lt;sup>a</sup> Mok T. Novel therapies [part of mini-symposium M12]. Presented at the 14th World Conference on Lung Cancer; Amsterdam, Netherlands; July 3-7, 2011.

ORR = objective response rate; PFS = progression-free survival; HR = hazard ratio; OS = overall survival; OR = odds ratio.

bind despite the T790M mutation, but can also bind to other HER receptors. These drugs can undoubtedly benefit patients failed by first-generation TKI, but whether T790M binding is responsible remains uncertain. The LUX-Lung series of trials with afatinib are illustrative; LUX-Lung 1 randomized patients who had received prior platinum chemotherapy and who had progressed after 12 or more weeks on erlotinib or gefitinib to either afatinib or placebo. The PFS, ORR, and symptom control outcomes strongly favoured afatinib, but os was not significantly different (79% of patients on the placebo arm received further lines of treatment). The large single-arm phase II LUX-Lung 2 trial included EGFR M+ patients at either first or second line. The ORR was 60%, but the PFS was an impressive 14 months. Results of LUX-Lung 3, which is now accrued and which randomized EGFR M+ patients in the first line to a fatinib or cisplatin-pemetrexed, are imminent and could lead to regulatory application.

Other resistance mechanisms to TKI in M+ patients include *MET* amplification (5%–20%) and, occasionally, epithelial–mesenchymal transition and

even transformation to a small-cell phenotype <sup>38</sup>. However, progressive disease on a first-generation TKI according to the formal Response Evaluation Criteria in Solid Tumors does not necessarily mean exhausted utility, because abrupt TKI cessation can, in about 20% of patients, induce a significant "flare" phenomenon that responds to immediate re-introduction of the same TKI <sup>39</sup>. Also, rechallenge with the same TKI after a "holiday" (during which chemotherapy may be given) is increasingly recognized as valuable <sup>40,41</sup>. There is an unmet need for biomarkers to guide the management of patients who experience technical progressive disease in front-line TKI, and there is evidence that resistance may differentially affect some metastases and not others—that is, clonal metastasis <sup>42,a</sup>.

Updated IPASS biomarker analysis <sup>32</sup> clearly showed that measurement of *EGFR* gene copy number

<sup>&</sup>lt;sup>a</sup> Zhong WZ. Genomic heterogeneity between primary tumor and its metastases. Presented at the 3rd International Thoracic Oncology Congress Dresden; Dresden, Germany; September 13–15, 2012.

by fluorescence *in-situ* hybridization (FISH) or of EGFR expression by immunohistochemistry (IHC) does not substitute for a mutation test. However, high copy number or IHC expression seems to be a weak surrogate for *EGFR* mutation positivity.

The NCIC BR.21 trial enrolled second- or third-line metastatic NSCLC patients who had exhausted their chemotherapy options. It showed an os benefit for erlotinib compared with placebo. A limited biomarker analysis suggested that high *EGFR* copy number by FISH (because of either gene amplification or high polysomy) <sup>43</sup> predicted a higher ORR (21% vs. 5%) and an improved os benefit from erlotinib (HR: 0.43 vs. 0.80 in FISH-negative patients). The FISH-positive control subjects had the worst os, but the most benefit from erlotinib, and compared with mutational status, FISH seemed to influence os more <sup>44</sup>.

Erlotinib was administered to more than 7000 patients in the large, open-label TRUST study (0–2 prior chemotherapies), with the German centres reporting their biomarker data. *EGFR* mutations and FISH positivity predicted response. Positivity by FISH also predicted PFS and OS. The EGFR IHC positivity weakly correlated with PFS and OS. Interestingly, 22% of patients were both IHC-positive and FISH-positive; about half to two thirds were IHC-positive, but FISH-negative; and 11%–21% were IHC-negative and FISH-negative, independent of histology <sup>45</sup>.

In nonrandomized studies such as TRUST, and even in randomized studies not using a placebo control, it is impossible to disentangle prognostic and predictive factors for PFS and OS; in this respect, BR.21 is highly valuable—as is ISEL46,47, a similar study that compared gefitinib with placebo, but in a more refractory population. In ISEL, which showed a nonsignificant benefit for gefitinib compared with placebo, high EGFR copy number predicted an os treatment effect (HR: 0.61 compared with placebo). An interaction test was significant (p = 0.045), indicating a genuinely different effect by copy number. The same applied to IHC status. EGFR mutations substantially predicted response (37.5% vs. 2.6%), but the data were too few to adjudicate survival effects. Results in the ISEL placebo group also suggested that FISH positivity was an adverse prognostic indicator (median survival time: 4.5 months vs. 6.4 months; HR: 1.41).

The BR.21, TRUST, and ISEL trials seem to imply utility for FISH and IHC as well as for *EGFR* mutational status, especially in Caucasian patients, in whom FISH positivity is more common than is mutation in unselected patients. In ISEL, 30.8% were FISH-positive and 12.1% were M+. Of the entire population, 20.2% were East Asian. In BR.21 (only 12% East Asian), 38% were FISH-positive and 18% were *EGFR* M+.

Ellis *et al.* performed a meta-analysis on the BR.21 and SATURN trials, two post-first-line trials, each with a placebo arm. Those authors concluded that EGFR IHC positivity is prognostic (weakly) for longer PFS and os, that *EGFR* FISH status was not prognostic, and that

EGFR mutations may be prognostic for os (perhaps confounded by crossover). Neither IHC nor FISH were recommended for "routine" prediction of erlotinib sensitivity; mutation positivity implied a better PFS on erlotinib, but mutation negativity did not preclude a benefit, and therefore EGFR mutation testing was not valuable after the first line <sup>48</sup>. In that analysis, some results for IHC, FISH (especially), and mutation status appeared to be discrepant between BR.21 and SATURN. In particular, FISH positivity was both predictive and negatively prognostic in BR.21, but not in SATURN. Notably, the BR.21 and SATURN patient populations were dissimilar.

The utility of FISH in the context of EGFR-TKI, especially in *EGFR* WT patients of any histology, should not be discounted for both prognosis and prediction.

Anti-EGFR monoclonal antibodies, especially cetuximab, added to chemotherapy in metastatic NSCLC generally produce modestly positive results. The FLEX study considered the addition of cetuximab to cisplatin-vinorelbine in EGFR IHC-positive metastatic NSCLC. Median os was increased by 1.2 months (HR: 0.871; p = 0.044)<sup>49</sup>. However, application of a scoring system ("H-score," continuous scale 0-300) revealed that 31% scored high (>200) and that the high-scoring patients (either histology) monopolized the os benefit (9.6 months vs. 12.0 months; HR: 0.73; p = 0.01). The low-score HR was 0.99. The interaction test was significant (p = 0.044)<sup>50</sup>. The Southwest Oncology Group 0819 study is attempting to prospectively confirm that result with cetux imab and carboplatin paclitaxel-bevacizumab.

Technical aspects of *EGFR* testing are beyond our scope <sup>51</sup>; however, microdissection and sequencing (Figure 1) may represent the current clinical standard. Allele-specific amplification—for example, Scorpions ARMS (DxS Limited, Manchester, U.K.)—is an alternative. Experimental mutation-specific antibodies are highly specific (97%–100%) and moderately sensitive (74.2%–100%) <sup>52–55</sup>. Detection of mutations in circulating tumour cells <sup>56,57</sup> or even circulating DNA <sup>58,59</sup> is rapidly being perfected.

In Canada (Table III), *EGFR* testing has been centralized in 5 laboratories, which might use different methodologies (for example, restriction fragment length polymorphism analysis, sequencing; Figure 1) and for which a minimum of five 5-µm sections are required, each with more than 100 tumour cells per section, and with the tumour cells representing more than 25% of the nucleated cells. Specimens are preferably microdissected and are better derived from core biopsies, although cell blocks and generous fine-needle aspirates may be adequate. Neither IHC nor FISH are routinely obtained.

# 3. ALK

As a driver oncogene, ALK (the anaplastic lymphoma kinase gene) was initially discovered in a

TABLE III Testing for *EGFR* mutation and selected uses of epidermal growth factor receptor inhibitors by province in Canada

Province	EGFR testing	First-line gefitinib (EGFR M+)	Maintenance erlotinib <sup>a</sup>
BC	√	√	√
AB	$\checkmark$	$\checkmark$	X
SK	$AZ^b$	Case by case	X
MB	AZ	Case by case	X
ON	AZ	$\checkmark$	X
QC	$\sqrt{}$	$\sqrt{}$	X
NS	AZ	X	X
NL	AZ	X	X
PE	AZ	X	X

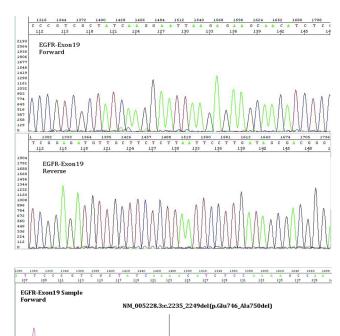
- a Not restricted by EGFR status.
- b Paid for by pharmaceutical company.

AZ = AstraZeneca Canada.

chromosomal rearrangement in anaplastic large-cell lymphoma 60. In 2007, Soda et al. described ALK activation in a subset of NSCLC that exhibited a "small inversion" in chromosome 2, fusing the normally separated EML4 (echinoderm microtubule-associated protein-like 4 gene) with ALK<sup>61</sup>. This EML4-ALK fusion transcript was detected in 5 of 75 Japanese NSCLC patients and in none of 261 patients with "other" cancers. Interestingly, although some EGFR (and KRAS) mutations were also found in the NSCLC cohort, none overlapped with the patients positive for EML4–ALK. The oncogenicity of the transcript was confirmed by transfection of expression plasmids into 3T3 cells, transforming them and subsequently showing tumorigenicity in nude mice. Although variants of the fusion transcript have been identified, in each case oncogenicity requires intact kinase function of ALK.

It was soon revealed that *EML4–ALK* lung carcinogenesis extended beyond Asia, characteristically occurring in middle-aged patients, usually neversmokers of either sex, and presenting as adenocarcinoma, especially the acinar histology in East Asia or the signet-ring or cribriform morphology in the West. This variant is always positive for TTF-1 <sup>62,63</sup>. Furthermore, mutual exclusivity between *EML4–ALK* and *EGFR* and *KRAS* mutations has been confirmed <sup>64</sup>.

The interest in *EML4–ALK* that has elevated its importance above its 2.5% incidence in NSCLC is its relatively specific and well-tolerated inhibitor, crizotinib. Crizotinib, originally in development as a Met inhibitor <sup>65</sup>, is also a potent Alk inhibitor. Entering human studies in 2006, the maximum tolerated dose was established as 250 mg twice daily. While that trial was open, the Morris *et al.* study was published, and the first *EML4–ALK* patient enrolled (receiving 300 mg orally, twice daily) enjoyed a rapid and dramatic response. Subsequently, intensive efforts were made to



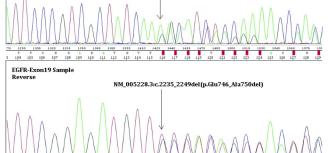


FIGURE 1 The DNA extracted from the macrodissected tissue specimen was amplified using primers specific for exon 19 of the EGFR gene. The polymerase chain reaction product was then purified and sequenced in both directions using the BigDye Terminator v3.1 Cycle Sequencing Kit (Applied Biosystems, Foster City, CA, U.S.A.). The sequences obtained were then compared with the U.S. National Center for Biotechnology Information reference sequence (EGFR: NM\_005228.3). In this case, a deletion is observed to span nucleotides 2235–2249, which results in the deletion of amino acids at positions 746–750 (inclusive) in exon 19 of the EGFR gene.

recruit NSCLC patients based on *ALK* rearrangements, and a high ORR was confirmed (10 responders in the first 19 patients), as reported in 2009 <sup>66</sup>. A further trial in 82 *ALK*-rearranged NSCLC patients appeared in 2010, showing a 57% ORR and 33% of patients with stable disease <sup>67</sup>. Further trials reported in 2011 that involved 119 (A8081001) and 136 (PROFILE 1005) patients led to conditional approval of crizotinib (Xalkori: Pfizer, Mission, KS, U.S.A.) by the U.S. Food and Drug Administration (FDA) <sup>67,68</sup> and by Health Canada more recently.

In those 255 patients (median age: 51 years; 48% men; 63% Caucasian, 30% Asian; 70% never-smokers, 28% former smokers; 96.5% with adenocarcinoma),

the orrs were 61% (A8081001) and 50% (Profile  $1005^b$ ). The PFS is expected to be  $\pm$  10 months (A8081001), with the os still uncertain. In patients progressing on the chemotherapy arm of the randomized second-line trial of pemetrexed or docetaxel compared with crizotinib (Profile 1007), Profile 1005 confirmed a very high orr<sup>b</sup>. An ongoing phase III trial, Profile 1014, is investigating first-line crizotinib compared with platin—pemetrexed.

The best detection method for *ALK* rearrangements in NSCLC is debatable. The current clinical standard the Break Apart FISH Probe kit (Abbott Molecular, Abbott Park, IL, U.S.A.), Figure 2—uses fluorescent green (5') and red (3') signals on loci in chromosome 2. normally so close together that they may fuse visually. Positivity consists of separation of these two markers by more than 2 signal diameters, or a red signal alone, in more than 15% tumour cells, counting more than 50 tumour cells. This is the companion assay approved by the FDA with crizotinib. Suitable for formalin-fixed, paraffin-embedded specimens, it is technically demanding and expensive, encouraging development of alternative methodologies, for example, reverse-transcriptase polymerase chain reaction (requiring knowledge of known fusion variants), DNA sequencing, or IHC. Immunohistochemistry, potentially with augmentation, may become a standard-of-care, high concordance with FISH having been established for IHC 3+ or IHC  $0^{69}$ . Intermediate IHC scores may, however, still require FISH. Several different antibodies are in development <sup>64</sup>.

Already, a crizotinib resistance mechanism has been identified, a "gatekeeper" mutation L1196M <sup>70–72</sup>. The gene *ROS* is also a target of crizotinib. Activation of *ROS* can be found in about 1.7% of NSCLC and can be assayed for; crizotinib appears to have marked activity in these cases <sup>73</sup>.

Pemetrexed may have exceptional activity in *ALK*-rearranged NSCLC <sup>74</sup>, with a response (in monotherapy or in combination with a platin) of 42% and a PFS of 9 months. Other publications have appeared in support <sup>75,76</sup>, but more recently, those findings have been questioned. The ongoing PROFILE studies should be informative.

#### 4. KRAS

The *ras* oncogenes were identified as cellular homologues of the Harvey and Kirsten strains of a mouse sarcoma virus <sup>77</sup>. Normally, Ras functions in signal transduction downstream of transmembrane receptor tyrosine kinases, especially EGFR, to which it is recruited by adaptor molecules, after binding of growth factors such as EGF and transforming growth

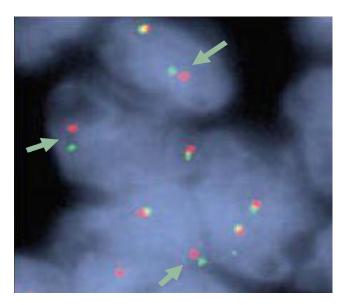


FIGURE 2 An example of a positive test with the Break Apart FISH Probe kit (Abbott Molecular, Abbott Park, IL, U.S.A.). A red and a green probe are hybridized to regions flanking the ALK translocation breakpoint; these probes will be separated by an intervening fusion of a translocated fragment (for example, EML4). That intervening fusion can be clearly seen here in several cells (arrows). In the other (normal) allele, the red and green probes are not separated, even appearing yellow (an artefact of visual overlap). The assay is designated positive if 15% or more of 50 or more cell nuclei demonstrate the split signal or an isolated red signal.

factor α to the receptor tyrosine kinase. Activation of Ras occurs through GTP binding, and as an intrinsic GTPase, it catalyzes GTP breakdown, enabling (then switching off) downstream signalling predominantly via the Raf/Mek/Erk downstream signal transduction pathway (the classical MAPK pathway). Erk activates transcription of genes mediating mitosis (and cell survival). At least 9 other pathways may be stimulated by Ras, including PI3K/Akt, a survival pathway <sup>78,79</sup>.

Of the *KRAS* mutations in NSCLC, 97% occur in exon 2, codon 12 or 13 80. These missense mutations impair the functionality of *ras* GTPase, locking the Ras signalling in active mode. Paradoxically, although the mutations inactivate Ras, the result is persistent signal activation. That persistence is one reason that Ras has been difficult to "drug"; it requires reactivation, not inactivation, to switch the signalling off.

Mutations of *ras* in NSCLC occur predominantly in "smoking adenocarcinoma" patients (30%–40%, Table I). In those patients, the mutations are G-to-T or G-to-C transversions (that is, pyrimidine swapped for purine); recently however, in never-smokers with adenocarcinoma, "transition" mutations [G to A (purine for purine)] have occasionally been found (approximately 15%), also probably oncogenic <sup>81</sup>. The mutation subtype may

b Riely GJ, Kim DW, Crino L, et al. Phase 2 data for crizotinib (PF-02341066) in ALK-positive advanced non-small cell lung cancer (NSCLC): PROFILE 1005 [abstract 1618]. Presented at the 14th World Conference on Lung Cancer; Amsterdam, Netherlands; July 3–7, 2011.

alter downstream signal activation, with potential implications <sup>82</sup> for prognosis. This heterogeneity may explain some of the conflicting data that characterize *KRAS* clinical research.

Currently, *KRAS* itself remains undruggable despite decades of effort. Attention has recently focused on inhibition of the Ras-contingent downstream signalling (especially Raf and Erk) or exploitation of synthetic lethality <sup>83</sup>.

Whether *KRAS* mutations influence EGFR-TKI responsiveness is contentious, and current Canadian recommendations discourage *ras* testing <sup>48</sup>. Studies indicating no benefit <sup>44,84</sup> have to be balanced by studies indicating that *KRAS* mutations are compatible with some benefit in, for example, maintenance—as with SATURN <sup>85</sup>. However, *KRAS* mutations may indicate a short PFS in the control arm and may therefore be adversely prognostic regardless of treatment.

The negative predictive effect of *KRAS* for treatment with anti-EGFR antibodies in colorectal cancer does not carry over to NSCLC treated with cetuximab; consider FLEX, for example, in which *KRAS* mutations were neither predictive nor prognostic <sup>86</sup>. However, *KRAS* mutations may sensitize tumours to antifolates such as pemetrexed <sup>74,87</sup>, possibly by upregulation of mir-181c, a micro RNA that can downregulate *KRAS*. Those observations require confirmation, given the high frequency of *KRAS* mutations in adenocarcinoma associated with smoking.

Currently, the chief value of *KRAS* lies in providing information about the other biomarkers that are directly druggable—that is, *EGFR* and *ALK*. The presence of mutated *KRAS* rules out *ALK* and *EGFR*, and *KRAS* may therefore form part of an efficient pathway in a testing algorithm.

#### 5. *MET*

Met is a receptor tyrosine kinase often expressed in epithelium. Its paracrine ligand, hepatocyte growth factor ("scatter factor"), is produced by stromal cells. Met signals via Ras, pi3κ/Akt, and stat, affecting mitosis, survival, angiogenesis, migration, invasion, and as implied, mesenchymal–epithelial transversion. Upregulation in cancer cells results in "invasive growth" <sup>88</sup>. Amplification of *MET* is documented in 4.1% of North American lung adenocarcinomas, but *MET* overexpression maybe more common<sup>c</sup>. Mutations in *MET* occur rarely.

Upregulation of MET may depend on prior exposure to therapy and may mediate resistance to it. Several studies indicate that MET amplification is responsible for  $\pm 20\%$  of resistance to EGFR-TKI <sup>89–92</sup>, prompting the

development of Met-inhibitory strategies. Tivantinib (ARO 197) is currently in phase III trial (MARQUEE) based on a successful randomized phase II study (erlotinib ± tivantinib). Non-squamous and KRAS M+ patients benefited most. MetMAb (Hoffmann-La Roche, Mississauga, ON), an anti-Met monoclonal antibody, achieved significant PFS and os benefit in a randomized phase II trial (OAM 4558g) with a similar "erlotinib ± experimental drug" design, but only in high expressors of MET (Met IHC 2+ or 3+). Detection by IHC (that is, expression) may be more reliable than detection by FISH (that is, amplification) in predicting MetMAb benefit. The effect in low expressors of Met appeared actually harmful, highlighting the importance of a companion diagnostic as MetMAb proceeds into phase III.

Crizotinib, although approved for *ALK*-rearranged metastatic NSCLC, is also a good Met inhibitor. An anecdotal report <sup>93</sup> of a rapid, durable response to crizotinib in a *MET*-amplified NSCLC patient with normal *ALK*, suggests that crizotinib may be suitable for that situation as well as for *ALK* rearrangements, as already shown for other types of cancer with *MET* amplification <sup>94,95</sup>.

MET will likely be the next major biomarker in metastatic NSCLC, given the speed with which the foregoing drugs (and others) <sup>88</sup> are approaching the clinic. How best to integrate them into the increasingly complex metastatic NSCLC algorithm will require substantial investment, but will likely pay major dividends.

# 6. SUMMARY

EGFR and ALK are biomarkers of current relevance in the management of non-squamous metastatic NSCLC and definitely predict a higher likelihood of benefit from EGFR-TKI and crizotinib respectively. Across Canada, efforts to promote access to testing require intensification. KRAS testing remains controversial—but interesting in the research setting and in testing algorithms as an efficiency tactic, because KRAS mutations are common and almost entirely rule out EGFR mutations and ALK rearrangements. MET amplification—or more likely, Met IHC—is required to optimize the development and clinical deployment of Met-directed therapies. Subject to confirmation, EGFR IHC ("H-score") might allow for the selection of patients benefiting from anti-EGFR monoclonal antibodies such as cetuximab.

The problem of inconsistent access to adequate tissue remains an important obstacle to the evolution of personalized medicine in metastatic NSCLC. The solution lies partly in the ongoing development of serum-based molecular assays, but for now, it lies in the education of interventional radiologists, thoracic surgeons, and respirologists, because optimal treatment of metastatic NSCLC is highly contingent on an adequate biopsy.

Varella–Garcia M, Iafrate J, Pao W, et al. ALK fusion and MET amplification as molecular biomarkers and therapeutic targets in advanced lung adenocarcinomas in the Lung Cancer Mutation Consortium [abstract 1348]. Presented at the 14th World Conference on Lung Cancer; Amsterdam, Netherlands; July 3–7, 2011.

#### 7. CONFLICT OF INTEREST DISCLOSURES

The authors declare consultancy work for Lilly and Roche; work on advisory boards for Pfizer, AstraZeneca, and Roche; membership in a speakers' bureau for Lilly.

# 8. REFERENCES

- Cheng L, Alexander RE, Maclennan GT, et al. Molecular pathology of lung cancer: key to personalized medicine. Mod Pathol 2012;25:347–69.
- Pao W, Miller V, Zakowski M, et al. EGF receptor gene mutations are common in lung cancers from "never smokers" and are associated with sensitivity of tumours to gefitinib and erlotinib. Proc Natl Acad Sci USA 2004;101:13306–11.
- 3. Aung KL, Board RE, Ellison G, *et al.* Current status and future potential of somatic mutation testing from circulating free DNA in patients with solid tumours. *HUGO J* 2010;4:11–21.
- Goto K, Ichinose Y, Ohe Y, et al. Epidermal growth factor receptor mutation status in circulating free DNA in serum: from IPASS, a phase III study of gefitinib or carboplatin/paclitaxel in non-small cell lung cancer. J Thorac Oncol 2012;7:115–21.
- 5. Cohen S. Isolation of a mouse submaxillary gland protein accelerating incisor eruption and eyelid opening in the newborn animal. *J Biol Chem* 1962;237:1555–62.
- Hollenberg MD, Cuatrecasas P. Epidermal growth factor: receptors in human fibroblasts and modulation of action by cholera toxin. *Proc Natl Acad Sci USA* 1973;70:2964–8.
- 7. Burgess AW. *EGFR* family: structure physiology signalling and therapeutic targets. *Growth Factors* 2008;26:263–74.
- Avraham R, Yarden Y. Feedback regulation of EGFR signalling: decision making by early and delayed loops. Nat Rev Mol Cell Biol 2011;12:104–17.
- 9. Ullrich A, Coussens L, Hayflick JS, *et al*. Human epidermal growth factor receptor CDNA sequence and aberrant expression of the amplified gene in A431 epidermoid carcinoma cells. *Nature* 1984;309:418–25.
- Mendelsohn J, Baselga J. Status of epidermal growth factor receptor antagonists in the biology and treatment of cancer. J Clin Oncol 2003;21:2787–99.
- Shigematsu H, Lin L, Takahashi T, et al. Clinical and biological features associated with epidermal growth factor receptor gene mutations in lung cancers. J Natl Cancer Inst 2005;97:339–46.
- 12. Tam IY, Chung LP, Suen WS, *et al.* Distinct epidermal growth factor receptor and *KRAS* mutation patterns in non-small cell lung cancer patients with different tobacco exposure and clinicopathologic features. *Clin Cancer Res* 2006;12:1647–53.
- 13. Bae NC, Chae MH, Lee MH, *et al. EGFR*, *ERBB2*, and *KRAS* mutations in Korean non-small cell lung cancer patients. *Cancer Genet Cytogenet* 2007;173:107–13.
- 14. Sordella R, Bell DW, Haber DA, Settleman J. Gefitinib-sensitizing *EGFR* mutations in lung cancer activate anti-apoptotic pathways. *Science* 2004;305:1163–7.
- 15. Takeuchi K, Ito F. EGF receptor in relation to tumor development: molecular basis of responsiveness of cancer cells to EGFR-targeting tyrosine kinase inhibitors. *FEBS J* 2010;277:316–26.

- Lynch TJ, Bell DW, Sordella R, et al. Activating mutations in the epidermal growth factor receptor underlying responsiveness of non-small-cell lung cancer to gefitinib. N Engl J Med 2004:350:2129–39.
- Paez JG, Jänne PA, Lee JC, et al. EGFR mutations in lung cancer: correlation with clinical response to gefitinib therapy. Science 2004;304:1497–500.
- Okamoto I. Epidermal growth factor receptor in relation to tumor development: EGFR-targeted anticancer therapy. FEBS J 2010;277:309–15.
- 19. Inoue A, Suzuki T, Fukuhara T, *et al.* Prospective phase II study of gefitinib for chemotherapy-naive patients with advanced non-small-cell lung cancer with epidermal growth factor receptor gene mutations. *J Clin Oncol* 2006;24:3340–6.
- Asahina H, Yamazaki K, Kinoshita I, et al. A phase II trial of gefitinib as first-line therapy for advanced non-small cell lung cancer with epidermal growth factor receptor mutations. Br J Cancer 2006;95:998–1004.
- Sutani A, Nagai Y, Udagawa K, et al. Gefitinib for non-smallcell lung cancer patients with epidermal growth factor receptor gene mutations screened by peptide nucleic acid-locked nucleic acid PCR clamp. Br J Cancer 2006;95:1483–9.
- Yoshida K, Yatabe Y, Park JY, et al. Prospective validation for prediction of gefitinib sensitivity by epidermal growth factor receptor gene mutation in patients with non-small cell lung cancer. J Thorac Oncol 2007;2:22–8.
- Sunaga N, Tomizawa Y, Yanagitani N, et al. Phase II prospective study of the efficacy of gefitinib for the treatment of stage III/IV non-small cell lung cancer with EGFR mutations, irrespective of previous chemotherapy. Lung Cancer 2007;56:383–9.
- 24. Tamura K, Okamoto I, Kashii T, et al. Multicentre prospective phase II trial of gefitinib for advanced non-small cell lung cancer with epidermal growth factor receptor mutations: results of the West Japan Thoracic Oncology Group trial (WJTOG 0403). Br J Cancer 2008;98:907–14.
- Sequist LV, Martins RG, Spigel D, et al. First-line gefitinib in patients with advanced non-small-cell lung cancer harboring somatic EGFR mutations. J Clin Oncol 2008;26:2442–9.
- Sugio K, Uramoto H, Onitsuka T, et al. Prospective phase II study of gefitinib in non-small cell lung cancer with epidermal growth factor receptor gene mutations. Lung Cancer 2009;64:314–18.
- 27. Rosell R, Moran T, Queralt C, *et al.* Screening for epidermal growth factor receptor mutations in lung cancer. *N Engl J Med* 2009;361:958–67.
- Lee JS, Park K, Kim SW, et al. A randomized phase III study of gefitinib versus standard chemotherapy (gemcitabine plus cisplatin) as a first-line treatment for never-smokers with advanced or metastatic adenocarcinoma of the lung [abstract PRS.4]. J Thorac Oncol 2009;4(suppl 1):S283.
- Ku GY, Haaland BA, de Lima Lopes G Jr. Gefitinib vs chemotherapy as first-line therapy in advanced non-small cell lung cancer: meta-analysis of phase III trials. *Lung Cancer* 2011;74:469–73.
- Maemondo M, Inoue A, Kobayashi K, et al. Gefitinib or chemotherapy for non-small-cell lung cancer with mutated EGFR. N Engl J Med 2010;362:2380–8.
- 31. Mitsudomi T, Morita S, Yatabe Y, *et al.* Gefitinib versus cisplatin plus docetaxel in patients with non-small-cell lung

- cancer harbouring mutations of the epidermal growth factor receptor (WJTOG 3405): an open label, randomised phase 3 trial. *Lancet Oncol* 2010;11:121–8.
- 32. Fukuoka M, Wu YL, Thongprasert S, *et al.* Biomarker analyses and final overall survival results from a phase III, randomized, open-label, first-line study of gefitinib versus carboplatin/paclitaxel in clinically selected patients with advanced non-small-cell lung cancer in Asia (IPASS). *J Clin Oncol* 2011;29:2866–74.
- 33. Zhou C, Wu YL, Chen G, *et al.* Erlotinib versus chemotherapy as first-line treatment for patients with advanced *EGFR* mutation–positive non-small-cell lung cancer (OPTIMAL, CTONG-0802): a multicenter, open-label, randomized, phase 3 study. *Lancet Oncol* 2011;12:735–42.
- 34. Rosell R, Carcereny E, Gervais R, *et al.* Erlotinib versus standard chemotherapy as first-line treatment for European patients with advanced *EGFR* mutation–positive non-small-cell lung cancer (ERUTAC): a multicenter, open-label, randomized phase 3 trial. *Lancet Oncol* 2012;13:239–46.
- Wu JY, Yu CJ, Shih JY, Yang CH, Yang PC. Influence of first-line chemotherapy and EGFR mutations on second-line gefitinib in advanced non-small cell lung cancer. Lung Cancer 2010;67:348–54.
- Cappuzzo F, Ciuleanu T, Stelmakh L, et al. Erlotinib as maintenance treatment in advanced non-small-cell lung cancer: a multicentre, randomized, placebo-controlled phase 3 study. Lancet Oncol 2010;11:521–9.
- 37. Yun CH, Mengwasser KE, Toms AV, *et al.* The T790M mutation in EGFR kinase causes drug resistance by increasing the affinity for ATP. *Proc Natl Acad Sci USA* 2008;105:2070–5.
- 38. Sequist LV, Waltman BA, Dias—Santagata D, *et al.* Genotypic and histological evolution of lung cancer acquiring resistance to EGFR inhibitors. *Sci Transl Med* 2011;3:75ra26.
- Chaft JE, Oxnard GR, Sima CS, Kris MG, Miller VA, Riely GJ. Disease flare after tyrosine kinase inhibitor discontinuation in patients with *EGFR*-mutant lung cancer and acquired resistance to erlotinib or gefitinib: implications for clinical trial design. *Clin Cancer Res* 2011;17:6298–303.
- Becker A, Crombag L, Heideman DA, et al. Retreatment with erlotinib: regain of TKI sensitivity following a drug holiday for patients with NSCLC who initially responded to EGFR-TKI treatment. Eur J Cancer 2011;47:2603–6.
- 41. Oxnard GR, Janjigian YY, Arcila ME, *et al.* Maintained sensitivity to EGFR tyrosine kinase inhibitors in *EGFR*-mutant lung cancer recurring after adjuvant erlotinib or gefitinib. *Clin Cancer Res* 2011;17:6322–8.
- 42. Park S, Holmes–Tisch AJ, Cho EY, *et al.* Discordance of molecular biomarkers associated with epidermal growth factor receptor pathway between primary tumors and lymph node metastasis in non-small cell lung cancer. *J Thorac Oncol* 2009;4:809–15.
- 43. Hirsch FR, Varella–Garcia M, Bunn PA Jr, *et al.* Molecular predictors of outcome with gefitinib in a phase III placebocontrolled study in advanced non-small-cell lung cancer. *J Clin Oncol* 2006;24:5034–42.
- 44. Zhu CQ, da Cunha Santos G, Ding K, *et al.* Role of *KRAS* and *EGFR* as biomarkers of response to erlotinib in National Cancer Institute of Canada Clinical Trials Group Study BR.21. *J Clin Oncol* 2008;26:4268–75.

- 45. Schneider CP, Heigener D, Schott-von-Römer K, et al. Epidermal growth factor receptor–related tumor markers and clinical outcomes with erlotinib in non-small cell lung cancer: an analysis of patients from German centers in the TRUST study. J Thorac Oncol 2008;3:1446–53.
- 46. Thatcher N, Chang A, Parikh P, Rodrigues Pereira J, et al. Gefitinib plus best supportive care in previously treated patients with refractory advanced non-small-cell lung cancer: results from a randomised, placebo-controlled, multicentre study (Iressa Survival Evaluation in Lung Cancer). Lancet 2005;366:1527–37.
- Hirsch FR, Varella–Garcia M, Bunn PA Jr, et al. Molecular predictors of outcome with gefitinib in a phase III placebocontrolled study in advanced non-small-cell lung cancer. J Clin Oncol 2006;24:5034–42.
- Ellis PM, Blais N, Soulieres D, et al. A systematic review and Canadian consensus recommendations on the use of biomarkers in the treatment of non-small cell lung cancer. J Thorac Oncol 2011;6:1379–91.
- 49. Pirker R, Pereira JR, Szczesna A, *et al.* Cetuximab plus chemotherapy in patients with advanced non-small-cell lung cancer (FLEX): an open-label randomised phase III trial. *Lancet* 2009;373:1525–31.
- Pirker R, Pereira JR, von Pawel J, et al. Epidermal growth factor receptor (EGFR) expression as a predictor for survival of first line chemotherapy plus cetuximab in FLEX study patients with advanced non-small cell lung cancer (NSCLC). Lancet Oncol 2012;13:33–42.
- Pirker R, Herth FJ, Kerr KM, et al. Consensus for EGFR mutation testing in non-small cell lung cancer: results from a European workshop. J Thorac Oncol 2010;5:1706–13.
- Yu J, Kane S, Wu J, et al. Mutation-specific antibodies for the detection of EGFR mutations in non-small-cell lung cancer. Clin Cancer Res 2009;15:3023–8.
- 53. Brevet M, Arcila M, Ladanyi M. Assessment of *EGFR* mutation status in lung adenocarcinoma by immunohistochemistry using antibodies specific to the two major forms of mutant *EGFR*. *J Mol Diagn* 2010;12:169–76.
- Kawahara A, Yamamoto C, Nakashima K, et al. Molecular diagnosis of activating EGFR mutations in non-small cell lung cancer using mutation-specific antibodies for immunohistochemical analysis. Clin Cancer Res 2010;16:3163–70.
- 55. Kato Y, Peled N, Wynes MW, et al. Novel epidermal growth factor receptor mutation-specific antibodies for non-small cell lung cancer: immunohistochemistry as a possible screening method for epidermal growth factor receptor mutations. J Thorac Oncol 2010;5:1551–8.
- Maheswaran S, Sequist LV, Nagrath S, et al. Detection of mutations in EGFR in circulating lung-cancer cells. N Engl J Med 2008;359:366–77.
- Sequist LV, Nagrath S, Toner M, Haber DA, Lynch TJ. The CTC-chip: an exciting new tool to detect circulating tumor cells in lung cancer patients. *J Thorac Oncol* 2009;4:281–3.
- 58. Yung TK, Chan KC, Mok TS, Tong J, To KF, Lo YM. Single-molecule detection of epidermal growth factor receptor mutations in plasma by microfluidics digital PCR in non-small cell lung cancer patients. *Clin Cancer Res* 2009;15:2076–84.
- Bai H, Mao L, Wang HS, et al. Epidermal growth factor receptor mutations in plasma DNA samples predict tumor response

- in Chinese patients with stages IIIB to IV non-small-cell lung cancer. *J Clin Oncol* 2009;27:2653–9.
- 60. Morris SW, Kirstein MN, Valentine MB, *et al.* Fusion of a kinase gene, *ALK*, to a nucleolar protein gene, *NPM*, in non-Hodgkin's lymphoma. *Science* 1994;263:1281–4.
- 61. Soda M, Choi YL, Enomoto M, *et al.* Identification of the transforming *EML4–ALK* fusion gene in non-small-cell lung cancer. *Nature* 2007;448:561–6.
- 62. Inamura K, Takeuchi K, Togashi Y, *et al. EML4–ALK* lung cancers are characterized by rare other mutations, a TTF-1 cell lineage, an acinar histology, and young onset. *Mod Pathol* 2009;22:508–15.
- Shaw AT, Yeap BY, Mino–Kenudson M, et al. Clinical features and outcome of patients with non-small-cell lung cancer who harbour EML4–ALK. J Clin Oncol 2009:27:4247–53.
- 64. Subramanian J, Corrales L, Soulieres D, Morgensztern D, Govindan R. Summary of presentations from the 46th Annual Meeting of the American Society of Clinical Oncology (2010) focus on tumor biology and biomarkers related to lung cancer. *J Thorac Oncol* 2011;6:399–403.
- 65. Ou SH. Crizotinib: a novel and first-in-class multitargeted tyrosine kinase inhibitor for the treatment of anaplastic lymphoma kinase rearranged non-small cell lung cancer and beyond. *Drug Des Devel Ther* 2011;5:471–85.
- 66. Kwak EL, Camidge DR, Clark J, et al. Clinical activity observed in a phase I dose escalation trial of an oral c-Met and Alk inhibitor, PF-02341066 [abstract 3509]. J Clin Oncol 2009;27:. [Available online at: http://www.asco.org/ASCOv2/Meetings/Abstracts?&vmview=abst\_detail\_view&confID=65&abstractID=30947; cited May 23, 2012]
- Kwak EL, Bang YJ, Camidge DR, et al. Anaplastic lymphoma kinase inhibition in non-small-cell lung cancer. N Engl J Med 2010;363:1693–703.
- 68. Crino L, Kim DW, Riely GJ, et al. Initial phase II results with crizotinib in advanced ALK-positive non-small cell lung cancer (NSCLC): PROFILE 1005 [abstract 7514]. J Clin Oncol 2011;29:. [Available online at: http://www.asco.org/ASCOv2/Meetings/Abstracts?&vmview=abst\_detail\_view&confID=102&abstractID=81844; cited May 23, 2012]
- Yang P, Kulig K, Boland JM, et al. Worse disease-free survival in never-smokers with ALK+ lung adenocarcinoma. J Thorac Oncol 2012;7:90–7.
- Choi YL, Soda M, Yamashita Y, et al. EML4–ALK mutations in lung cancer that confer resistance to Alk inhibitors. N Engl J Med 2010;363:1734–9.
- Katayama R, Khan TM, Benes C, et al. Therapeutic strategies to overcome crizotinib resistance in non-small cell lung cancers harboring the fusion oncogene EML4–ALK. Proc Natl Acad Sci USA 2011;108:7535–40.
- 72. Sakamoto H, Tsukaguchi T, Hiroshima S, *et al.* CH5424802, a selective Alk inhibitor capable of blocking the resistant gatekeeper mutant. *Cancer Cell* 2011;19:679–90.
- Bergethon K, Shaw AT, Ou SH, et al. ROS1 rearrangements define a unique molecular class of lung cancers. J Clin Oncol 2012;30:863–70.
- 74. Camidge DR, Kono SA, Lu X, *et al.* Anaplastic lymphoma kinase gene rearrangements in non-small cell lung cancer are associated with prolonged progression-free survival on pemetrexed. *J Thorac Oncol* 2011;6:774–80.

- 75. Takeda M, Okamoto I, Sakai K, *et al.* Successful long-term treatment with pemetrexed of NSCLC associated with *EML4–ALK* and low thymidylate synthase expression. *Clin Lung Cancer* 2012;13:157–9.
- Lee JO, Kim TM, Lee SH, et al. Anaplastic lymphoma kinase translocation: a predictive biomarker of pemetrexed in patients with non-small cell lung cancer. J Thorac Oncol 2011;6:1474–80.
- Chang EH, Gonda MA, Ellis RW, Scolnick EM, Lowy DR. Human genome contains four genes homologous to transforming genes of Harvey and Kirsten murine sarcoma viruses. *Proc Natl Acad Sci USA* 1982;79:4848–52.
- 78. Vakiani E, Solit DB. *KRAS* and *BRAF*: drug targets and predictive biomarkers. *J Pathol* 2011;223:219–29.
- Xu N, Lao Y, Zhang Y, Gillespie DA. Akt: a double-edged sword in cell proliferation and genome stability. *J Oncol* 2012;2012:951724.
- 80. Forbes S, Clements J, Dawson E, *et al.* COSMIC 2005. *Br J Cancer* 2006;94:318–22.
- 81. Riely GJ, Kris MG, Rosenbaum D, *et al.* Frequency and distinctive spectrum of *KRAS* mutations in never smokers with lung adenocarcinoma. *Clin Cancer Res* 2008;14:5731–4.
- Ihle NT, Byers LA, Kim ES, et al. Effect of KRAS oncogene substitutions on protein behavior: implications for signaling and clinical outcome. J Natl Cancer Inst 2012;104:228–39.
- 83. Luo J, Emanuele MJ, Li D, *et al.* A genome-wide RNAi screen identifies multiple synthetic lethal interactions with the *ras* oncogene. *Cell* 2009;137:835–48.
- 84. Eberhard DA, Johnson BE, Amler LC, *et al.* Mutations in the epidermal growth factor receptor and in *KRAS* are predictive and prognostic indicators in patients with non-small-cell lung cancer treated with chemotherapy alone and in combination with erlotinib. *J Clin Oncol* 2005;23:5900–9.
- Brugger W, Triller N, Blasinska–Morawiec M, et al. Prospective molecular marker analyses of EGFR and KRAS from a randomized, placebo-controlled study of erlotinib maintenance therapy in advanced non-small-cell lung cancer. J Clin Oncol 2011;29:4113–20.
- O'Byrne KJ, Gatzemeier U, Bondarenko I, et al. Molecular biomarkers in non-small-cell lung cancer: a retrospective analysis of data from the phase 3 FLEX study. Lancet Oncol 2011;12:795–805.
- 87. Bacus S. *KRAS* mutation and amplification status predicts sensitivity to antifolate therapies in non-small cell lung cancer [abstract PR-2]. *Mol Cancer Ther* 2011;10 (suppl 1):.
- Feng Y, Thiagarajan PS, Ma PC. Met signaling: novel targeted inhibition and its clinical development in lung cancer. *J Thorac Oncol* 2012;7:459–67.
- Ma PC, Tretiakova MS, MacKinnon AC, et al. Expression and mutational analysis of MET in human solid cancers. Genes Chromosomes Cancer 2008;47:1025–37.
- Engelman JA, Zejnullahu K, Mitsudomi T, et al. MET amplification leads to gefitinib resistance in lung cancer by activating ErbB3 signaling. Science 2007;316:1039–43.
- 91. Cappuzzo F, Jänne PA, Skokan M, *et al. MET* increased gene copy number and primary resistance to gefitinib therapy in non-small-cell lung cancer patients. *Ann Oncol* 2009;20:298–304.
- Bean J, Brennan C, Shih JY, et al. MET amplification occurs with or without T790M mutations in EGFR mutant lung tumors

#### BIOMARKERS AFFECTING CLINICAL PRACTICE

- with acquired resistance to gefitinib or erlotinib. *Proc Natl Acad Sci USA* 2007;104:20932–7.
- 93. Ou SH, Kwak EL, Siwak–Tapp C, *et al.* Activity of crizotinib (PF02341066), a dual mesenchymal–epithelial transition (*MET*) and anaplastic lymphoma kinase (*ALK*) inhibitor, in a non-small cell lung cancer patient with *de novo MET* amplification. *J Thorac Oncol* 2011;6:942–6.
- 94. Lennerz JK, Kwak EL, Ackerman A, *et al. MET* amplification identifies a small and aggressive subgroup of esophagogastric adenocarcinoma with evidence of responsiveness to crizotinib. *J Clin Oncol* 2011;29:4803–10.
- 95. Chi A, Kwak EL, Clark JW, *et al.* Clinical improvement and rapid radiographic regression induced by a Met inhibitor in a patient with *MET*-amplified glioblastoma [abstract 2072].

J Clin Oncol 2011;29:. [Available online at: http://www.asco. org/ASCOv2/Meetings/Abstracts?&vmview=abst\_detail\_view&confID=102&abstractID=82407; cited May 23, 2012]

Correspondence to: Mark D. Vincent, 790 Commissioners Road East, London, Ontario N6A 4L6. E-mail: mark.vincent@lhsc.on.ca

- \* Department of Medical Oncology, London Regional Cancer Program, London Health Sciences Centre, London, ON.
- † Departments of Medical Oncology and Pathology, University Health Network, Princess Margaret Hospital-Ontario Cancer Institute, Toronto, ON.